

3. H. J. Zwally, J. C. Comiso, C. L. Parkinson, D. J. Cavalieri, P. Gloersen, *J. Geophys. Res.* **107**, 9.1 (2002).
4. A. B. Watkins, I. Simmonds, *J. Climate* **13**, 4441 (2000).
5. D. J. Cavalieri, P. Gloersen, C. L. Parkinson, J. C. Comiso, H. J. Zwally, *Science* **278**, 1104 (1997).
6. X. Wu, W. F. Budd, T. H. Jacka, *Ann. Glaciol.* **29**, 61 (1999).
7. S. Nicol *et al.*, *Nature* **406**, 504 (2001).
8. X. Yuan, D. G. Martinson, *J. Clim.* **13**, 1697 (2000).
9. P. R. Wilson *et al.*, *Mar. Ecol. Prog. Ser.* **213**, 301 (2001).
10. C. Barbraud, H. Weimerskirch, *Nature* **411**, 183 (2001).
11. M. A. J. Curran, G. B. Jones, *J. Geophys. Res.* **105**, 20451 (2000).
12. J. O'Dwyer *et al.*, *Geophys. Res. Lett.* **27**, 1159 (2000).
13. E. C. Pasteur, R. Mulvaney, D. A. Peel, E. S. Saltzman, P.-Y. Whung, *Ann. Glaciol.* **21**, 169 (1995).
14. K. A. Welch, P. A. Mayewski, S. I. Whitlow, *Geophys. Res. Lett.* **20**, 443 (1993).
15. V. Morgan *et al.*, *J. Glaciol.* **43**, 3 (1997).
16. M. A. J. Curran, T. D. van Ommen, V. Morgan, *Ann. Glaciol.* **27**, 385 (1998).
17. M. A. J. Curran *et al.*, *Ann. Glaciol.* **35**, 333 (2002).
18. These data are available on the Antarctic Cooperative Research Center and Australian Antarctic Division Climate Data Sets Web site, www.antcrc.utas.edu.au/~jacka/climate.html.
19. A. Minikin *et al.*, *J. Geophys. Res.* **103**, 10975 (2002).
20. M. Legrand, C. Feniet-Saigne, *Geophys. Res. Lett.* **18**, 187 (1991).
21. A. Turnipseed, A. Ravishankara, in *Dimethylsulphide: Oceans, Atmosphere, and Climate*, G. Restelli, G. Angeletti, Eds. (Kluwer Academic, Belgirate, Italy 1993), pp. 185–195.
22. L. K. Armand, A. Leventer, in *Sea Ice: An Introduction to its Physics, Chemistry, Biology, and Geology*, D. N. Thomas, G. S. Dieckmann, Eds. (Blackwell, Oxford, UK, 2003), pp. 333–372.
23. V. I. Morgan, I. D. Goodwin, D. M. Etheridge, C. W. Wookey, *Nature* **354**, 5066 (1991).
24. H. J. Zwally, C. L. Parkinson, J. C. Comiso, *Science* **220**, 1005 (1983).
25. E. J. Murphy, A. Clarke, C. Symon, J. Priddle, *Deep Sea Res.* **42**, 1045 (1995).
26. W. K. de la Mare, *Nature* **389**, 57 (1997).
27. C. L. Parkinson, *Ann. Glaciol.* **14**, 221 (1990).
28. S. Vaughan, *Polar Record* **36**, 345 (2000).
29. W. B. White, R. G. Peterson, *Nature* **380**, 699 (1996).
30. X. Yuan, D. G. Martinson, *Geophys. Res. Lett.* **28**, 3609 (2001).

Supporting Online Material

www.sciencemag.org/cgi/content/full/302/5648/1203/DC1
SOM Text
Fig. S1
References

11 June 2003; accepted 8 October 2003

Early Allelic Selection in Maize as Revealed by Ancient DNA

Viviane Jaenicke-Després,¹ Ed S. Buckler,² Bruce D. Smith,³
M. Thomas P. Gilbert,⁴ Alan Cooper,⁴ John Doebley,⁵
Svante Pääbo^{1*}

Maize was domesticated from teosinte, a wild grass, by ~6300 years ago in Mexico. After initial domestication, early farmers continued to select for advantageous morphological and biochemical traits in this important crop. However, the timing and sequence of character selection are, thus far, known only for morphological features discernible in corn cobs. We have analyzed three genes involved in the control of plant architecture, storage protein synthesis, and starch production from archaeological maize samples from Mexico and the southwestern United States. The results reveal that the alleles typical of contemporary maize were present in Mexican maize by 4400 years ago. However, as recently as 2000 years ago, allelic selection at one of the genes may not yet have been complete.

The wild grass, teosinte (*Zea mays* ssp. *parviglumis*), from which maize (*Zea mays* ssp. *mays*) was domesticated, is endemic to southern and western Mexico (1). The earliest undisputed archaeological evidence of domesticated maize is 6250 years old (2). However, recent molecular data suggest that domestication could have begun as early as 9000 years ago and that the Balsas River Valley in southern Mexico is the likely geographical origin of domestication (3). The early history of character selection in maize is documented in the archaeological record by morphological features discernible in cobs. For example, an increase in the number of rows of

kernels and a reduction in glume size have been noted in early maize cobs (4). By 5500 years ago, kernel size had also increased (5). However, nothing is currently known about when characters not observable from the morphology of cobs, such as plant architecture and starch properties, were selected by early farmers.

Recently, a number of genetic loci associated with phenotypic differences between maize and teosinte have been identified (6–9), and three genes involved in such differences have been cloned and relatively well characterized in function (7, 9, 10). In each of these genes, the allelic diversity in maize compared with teosinte has been shown to be reduced, presumably as a result of selection by early farmers. The first gene, *teosinte branched 1* (*tb1*), carries a maize variant that represses the growth in axillary meristems, leading to the unbranched plant architecture typical of maize. It also contributes to the presence of female cobs on the primary branches in maize rather than male tassels as in teosinte (11, 12). The second gene encodes the prolamins box binding factor (*pb1*), which is involved in the control of expression of

seed storage proteins in the kernel (13–15), whereas the third gene, *sugary 1* (*su1*), encodes a starch debranching enzyme expressed in kernels (16). Together with branching enzymes, this enzyme determines the structure of amylopectin (16, 17). The chain length of amylopectin, as well as the ratio of amylose to amylopectin, is important for the gelatinization properties of starch (9) and, thus, affects the textural properties of tortillas (18, 19).

Because DNA in archaeological remains is generally degraded to small sizes (20), we identified fragments in each gene that are short enough to allow amplification from ancient corn cobs yet distinguish between the spectrum of gene variants (alleles) found in present-day maize and teosinte (10). For *tb1*, the allelic variation in contemporary maize and teosinte is well described (10). This allowed us to choose a fragment of 56 base pairs (bp) for which maize carries a single allele, Tb1-M1; this allele has a frequency of 36% in teosinte, where a total of six additional alleles exist. In order to characterize the contemporary variation in *pb1* and *su1*, we sequenced a longer segment of each gene in 66 maize landraces from South, Middle, and North America as well as 23 teosinte *parviglumis* lines (10). The estimated number of alleles segregating in maize is reduced about threefold at *pb1* and *su1* in comparison to teosinte (fig. S1, B and C). At *pb1*, we selected a 25-bp fragment in which the alleles Pbf-M1 and Pbf-M2 are carried in 97% and 3% of maize, whereas the same alleles are carried in 17% and 83% of teosinte, respectively (Fig. 1). At *su1*, we selected a 60-bp fragment in which two major alleles, Su1-M1 and Su1-M2, are carried in maize at a frequency of 30 and 62%, respectively, whereas they both are carried in teosinte at a frequency of about 7% (Fig. 1; table S2) (10).

We investigated five maize cobs from the Ocampo Caves in northeastern Mexico (Fig. 2) and six cobs from Tularosa Cave in the Mogollon highlands in New Mexico (10).

¹Max Planck-Institute for Evolutionary Anthropology, Deutscher Platz 6, D-04103 Leipzig, Germany. ²United States Department of Agriculture/Agricultural Research Service and Department of Genetics, North Carolina State University, Raleigh, NC 27695, USA. ³Archaeobiology Program, Department of Anthropology, National Museum of Natural History, Smithsonian Institution, Washington, DC 20560, USA. ⁴Henry Wellcome Ancient Biomolecules Centre, Department of Zoology, University of Oxford, Oxford OX1 3PS, UK. ⁵Laboratory of Genetics, University of Wisconsin, Madison, WI 53706, USA.

*To whom correspondence should be addressed. E-mail: paabo@eva.mpg.de

Each cob was dated directly using tandem accelerator mass spectroscopy (10). Two cobs from the Ocampo Caves are around 4300 years old, whereas the other three are between 2300 and 2800 years old. Two cobs from Tularosa Cave are around 1900 years old, and the remaining four are between 650 and 900 years old (Fig. 3). From 140 to 200 mg of each cob, we extracted DNA and amplified the fragments containing the diagnostic nucleotide for the alleles at *tb1*, *pbf*, and *su1* (10). DNA sequences were reconstructed from multiple clones derived from several amplification products from each sample, according to established procedures (21), and were compared to modern maize and teosinte alleles (Fig. 3). To verify the results, samples from two cobs (NMNH246279, 314984 cob2) were sent to our Oxford laboratory where DNA was extracted and DNA sequences from all three genes were independently determined. In all cases, the results in Oxford were identical to those in our Leipzig laboratory (10).

At *tb1*, the allele Tb1-M1 common in contemporary maize was found in all 11 cobs analyzed. At *pbf*, all ancient samples analyzed carried the Pbf-M1 allele, which occurs in an estimated 97% of modern maize. At *su1*, all cobs from Mexico carried the allele Su1-M2. Among the cobs from the southwestern United

States, the four younger cobs carried the M1 allele, whereas the two 2000-year-old cobs were heterozygous. One of these cobs carried the alleles M1 and M2, and the other carried M1 and the allele Su1-T1, which has not been seen in contemporary maize but occurs at an estimated frequency of 4% in modern teosinte (Fig. 1). In the case of samples found to be homozygous, heterozygosity cannot be rigorously ruled out because low template copy numbers and primer failure due to unknown alleles are impossible to exclude.

The results show that alleles known to occur in modern maize at the genes *tb1*, *pbf*, and *su1* were already present in maize 4400 years ago in Mexico. This shows that plant morphology as well as biochemical properties of the protein and starch were selected early in the history of maize, and before maize was introduced to the southwestern United States.

It has been suggested that genes which make harvesting of grasses easier may have been selected before genes increasing yield (22). Because the allele Tb1-M1 at the *tb1* gene causes the female cobs to be attached close to the main axis of the plant, the presence of Tb1-M1 in maize 4400 years ago is consistent with this suggestion. However, the morphology of corn cobs, ranging from 6250 years old (2) to 5500 years old (5) to the 4400-year-old cobs from the Ocampo Caves

analyzed here (23) (Fig. 2), shows that cob size increased continuously during the first 2000 years of selection by early farmers. The fact that *pbf* and *su1*, involved in protein and starch quality, respectively, carried the alleles typical of modern maize 4400 years ago suggests that kernel quality along with cob size was an early target of selection. However, the lack of available early maize samples means it is not possible to know how complete the selection process was by 4400 to 4300 years ago. In fact, the observation that a 2000-year-old cob from New Mexico carries a *su1* allele which occurs today in teosinte but is very rare or absent in maize indicates that the selection process at *su1* was not complete at that time. It is interesting that this gene may influence the pasting properties of maize starch and that this property, which is of importance for the suitability of the corn for making tortillas, may thus have been selected long after the initial domestication of maize. Analyses of more maize samples are necessary to assess to what extent this is indeed the case.

Maize appears in the archaeological record of the southwestern United States by ~3200 years ago (24). It appears infrequently in the eastern United States from 2100 years ago onward, but it did not become a dominant crop there until ~1200 years ago (25). Today, the allele Su1-M1 is found only in inbred maize lines that have a substantial contribution from Northern Flint, one of the two parents of modern Corn Belt maize (9), whereas Su1-M2 is present in other maize inbred lines. The absence of Su1-M1 in the Mexican archaeological samples and the presence of Su1-M1 at a high frequency in New Mexico 1000 years ago (Fig. 1) suggests that this allele may be closely connected to maize that

Downloaded from <http://science.sciencemag.org/> on June 2, 2020

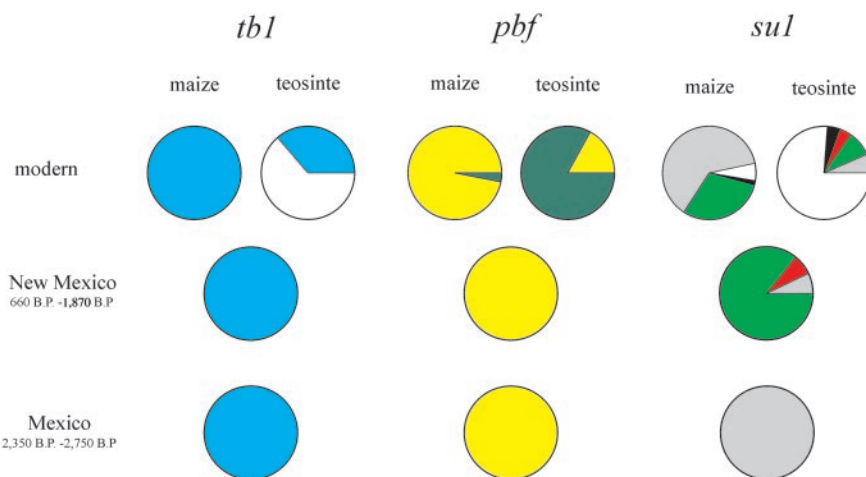


Fig. 1. Schematic illustration of the allelic diversity in modern maize, modern teosinte, and ancient maize from Mexico and New Mexico. Alleles shared between modern maize, teosinte, and/or ancient maize share the same color, whereas all other alleles are pooled in one white segment. For *tb1*, six nonshared alleles exist in teosinte; for *su1*, five nonshared alleles exist in maize and eight in teosinte. Blue, Tb1-M1; yellow, Pbf-M1; green, Su1-M1; gray, Su1-M2; red, Su1-T1; and black, Su1-M3. The areas of the segments approximate allele frequencies.

Fig. 2. A maize cob (248/E20; L2b/3) from the Ocampo Caves (Valenzuela cave), dated to 3890 ± 60 years before the present. Length, 47 mm.

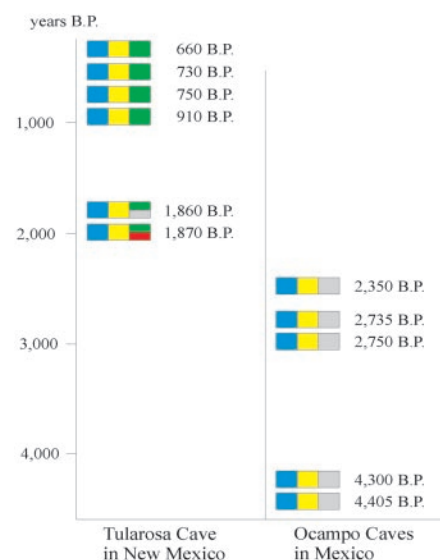
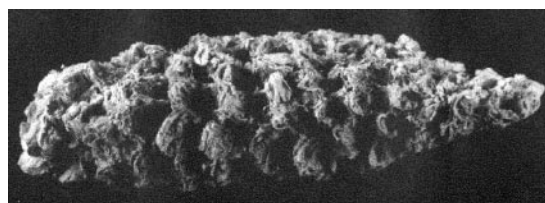


Fig. 3. Alleles in ancient cobs from New Mexico and Mexico. In every cob, three genes were analyzed: *tb1*, *pbf*, and *su1*. To the right of the cobs, their ages are given in calibrated years before present (B.P.). Colors are as in Fig. 1.

REPORTS

later became Northern Flint before it was introduced into the eastern United States. The fact that both 2000-year-old cobs were heterozygous for Su1-M1 may indicate that the predominance of Su1-M1 in North America was established sometime between 2000 and 1000 years ago in the southwestern United States. Analysis of maize remains from additional archaeological sites of this time period will be required to determine when Su1-M1 became predominant in the southwestern United States.

In conclusion, by 4400 years ago, early farmers had already had a substantial homogenizing effect on allelic diversity at three genes associated with maize morphology and biochemical properties of the corn cob. Thus, selection by farmers had profound genomic effects relatively early in the history of this crop. As more genes involved in selected features become identified in maize as well as other crops, the ability to determine nuclear gene sequences from domesticated plants recovered from archaeological excavations will make it possible to follow comprehensively the genetic consequences of domestication over time.

References and Notes

1. B. D. Smith, *The Emergence of Agriculture* (Scientific American Library, New York, 1998).
2. D. R. Piperno, K. V. Flannery, *Proc. Natl. Acad. Sci. U.S.A.* **98**, 2101 (2001).
3. Y. Matsuoka *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **99**, 6080 (2002).
4. B. Benz, *Proc. Natl. Acad. Sci. U.S.A.* **98**, 2104 (2001).
5. B. F. Benz, A. D. Long, *Curr. Anthropol.* **41**, 459 (2000).
6. J. Doebley, *Trends Genet.* **8**, 302 (1992).
7. R. L. Wang, A. Stec, J. Hey, L. Lukens, J. Doebley, *Nature* **398**, 236 (1999).
8. Y. Vigouroux *et al.*, *Proc. Natl. Acad. Sci. U.S.A.* **99**, 9650 (2002).
9. S. R. Whitt, L. M. Wilson, M. I. Tenaillon, B. S. Gaut, E. S. Buckler, *Proc. Natl. Acad. Sci. U.S.A.* **99**, 12959 (2002).
10. Material and Methods are available as supporting material on Science Online.
11. J. Doebley, A. Stec, L. Hubbard, *Nature* **386**, 485 (1997).
12. J. Doebley, A. Stec, C. Gustus, *Genetics* **141**, 333 (1995).
13. T. Ueda, Z. Wang, N. Pham, J. Messing, *Mol. Cell. Biol.* **14**, 4350 (1994).
14. Z. Wang, T. Ueda, J. Messing, *Gene* **223**, 321 (1998).
15. J. Vicente-Carbajosa, S. P. Moose, R. L. Parson, R. J. Schmidt, *Proc. Natl. Acad. Sci. U.S.A.* **94**, 7685 (1997).
16. A. Rahman, K. Wong, J. Jane, A. M. Myers, M. G. James, *Plant Physiol.* **117**, 425 (1998).
17. M. G. James, D. S. Robertson, A. M. Myers, *Plant Cell* **7**, 417 (1995).
18. J. Jane, J. Chen, *Cereal Chem.* **69**, 60 (1992).
19. J. Jane *et al.*, *Cereal Chem.* **76**, 629 (1999).
20. O. Handt, M. Höss, M. Krings, S. Pääbo, *Experientia* **50**, 524 (1994).
21. O. Handt, M. Krings, R. H. Ward, S. Pääbo, *Am. J. Hum. Genet.* **59**, 368 (1996).
22. E. S. Buckler, J. M. Thornsberry, S. Kresovich, *Genet. Res. Camb.* **77**, 213 (2001).
23. B. D. Smith, *Lat. Am. Antiq.* **8**, 342 (1997).
24. B. B. Huckell, *J. Prehist.* **10**, 305 (1996).
25. B. D. Smith, *Science* **246**, 1566 (1989).
26. We thank S. E. Nash, J. Haas, D. Martin-Ross, and G. Feinman, Field Museum; E. W. Wood, D. Pfister, and W. Wetterstrom, Harvard University Herbaria, for access to and permission to sample their archaeolog-

ical maize collections; B. Rauh for the *pbf* data; S. Whitt for the *su1* data; members of the Doebley Lab for preparing the modern DNAs; M. Przeworski, D. Serre, S. Ptak, M. Kuch, H. Poinar, G. Weiss, B. Wolter, and especially M. Hofreiter for helpful discussions; and the Bundesministerium für Bildung und Forschung, the Max Planck Gesellschaft, Wellcome Trust and the National Science Foundation for financial support.

Supporting Online Material

www.sciencemag.org/cgi/content/full/302/5648/1206/DC1

Materials and Methods

Figs. S1 and S2

Tables S1 and S2

References

10 July 2003; accepted 15 September 2003

Control of Nutrient-Sensitive Transcription Programs by the Unconventional Prefoldin URI

Matthias Gstaiger,^{1*} Brian Luke,² Daniel Hess,¹ Edward J. Oakeley,¹ Christiane Wirbelauer,¹ Marc Blondel,^{2,†} Marc Vigneron,³ Matthias Peter,² Wilhelm Krek^{1*‡}

Prefoldins (PFDs) are members of a recently identified, small-molecular weight protein family able to assemble into molecular chaperone complexes. Here we describe an unusually large member of this family, termed URI, that forms complexes with other small-molecular weight PFDs and with RPB5, a shared subunit of all three RNA polymerases. Functional analysis of the yeast and human orthologs of URI revealed that both are targets of nutrient signaling and participate in gene expression controlled by the TOR kinase. Thus, URI is a component of a signaling pathway that coordinates nutrient availability with gene expression.

The evolutionarily conserved phosphatidylinositolide (PI) 3-kinase-related kinase TOR (target of rapamycin) pathway occupies a central role in the integration and transduction of nutritional cues into a coherent cell-growth and proliferative response. Nutrient-rich conditions sustain TOR activity, which in turn fuels cell growth. In contrast, nutrient-depleted environments (or treatment with the immunosuppressant rapamycin) cause inhibition of TOR, which results in the activation of a response program that includes the induction of nutrient-sensitive gene expression (*1–4*). Here we describe an evolutionarily conserved member of the prefoldin (PFD) family, termed URI (for Unconventional prefoldin RPB5 Interactor), that participates in the regulation of nutrient-sensitive, TOR-dependent transcription programs.

While searching for proteins associated with the F-box protein SKP2 (for S-phase kinase-associated protein 2), which is the

substrate recognition component of the cell cycle-regulatory SCF^{SKP2} ubiquitin ligase (*5, 6*), we identified a member of the PFD family of small-molecular weight (14 to 23 kD) proteins. PFD family members are composed of N- and C-terminal, α -helical, coiled-coil structures connected by either one (β -class PFDs) or two (α -class PFDs) β hairpins (*7*). Yeast and human PFDs 1 to 6 assemble into an $\alpha 2\beta 4$ hexameric complex, referred to as the prefoldin/GimC complex, that functions as a molecular chaperone in actin and tubulin folding (*8–10*). Because the identified protein is an α -class PFD that associates with SKP2 in vivo, we termed it STAP1 (for SKP2-associated alpha PFD 1) (fig. S1).

We reasoned that STAP1, which is not part of prefoldin/GimC, could be a component of a unknown prefoldin-like complex. Mass spectrometric identification of STAP1-associated proteins from HeLa cells (Fig. 1A) revealed two β -class PFDs, PFD2 and PFD4-related (PFD4r); three proteins of unknown function; and four proteins whose functions have been linked to transcription. These are RPB5, a subunit shared by RNA polymerases (pols) I, II, and III (*11*); the adenosine triphosphatases TIP48 and TIP49 (*12, 13*); and RBP5-mediated protein (RMP), which is known to bind RPB5 (*14*). We refer to RMP hereafter as URI.

On gel filtration, STAP1 eluted with URI as a single peak at ~ 1 MD (fig. S2). RPB5, TIP48, and TIP49 were present in multiple fractions, including those that contained STAP1 and URI. A fraction of total

¹Friedrich Miescher Institut, Maulbeerstrasse 66, CH-4058 Basel, Switzerland. ²Eidgenössische Technische Hochschule (ETH)–Hönggerberg, Institute of Biochemistry, CH-8093 Zurich, Switzerland. ³Unité Mixte de Recherche 7100 CNRS–Université Louis Pasteur, Ecole Supérieure de Biotechnologie de Strasbourg, Boulevard Sébastien Brant, Boite postale 10413, 67412 Illkirch Cedex.

*Present address: ETH–Hönggerberg, Institute of Cell Biology, CH-8093 Zurich, Switzerland.

†Present address: Cell Cycle Group, Station Biologique du CNRS, Place Georges Teissier, F-29680 Roscoff, France.

‡To whom correspondence should be addressed. E-mail: wilhelm.krek@cell.biol.ethz.ch

Early Allelic Selection in Maize as Revealed by Ancient DNA

Viviane Jaenicke-Després, Ed S. Buckler, Bruce D. Smith, M. Thomas P. Gilbert, Alan Cooper, John Doebley and Svante Pääbo

Science **302** (5648), 1206-1208.
DOI: 10.1126/science.1089056

ARTICLE TOOLS	http://science.sciencemag.org/content/302/5648/1206
SUPPLEMENTARY MATERIALS	http://science.sciencemag.org/content/suppl/2003/11/13/302.5648.1206.DC1
RELATED CONTENT	http://science.sciencemag.org/content/sci/302/5648/1158.full
REFERENCES	This article cites 23 articles, 10 of which you can access for free http://science.sciencemag.org/content/302/5648/1206#BIBL
PERMISSIONS	http://www.sciencemag.org/help/reprints-and-permissions

Use of this article is subject to the [Terms of Service](#)

Science (print ISSN 0036-8075; online ISSN 1095-9203) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science* is a registered trademark of AAAS.

American Association for the Advancement of Science